

RESEARCH ARTICLE

Open Access

# The role of complement factor H in gestational diabetes mellitus and pregnancy



Junxian Li<sup>1†</sup>, Ying Shen<sup>2†</sup>, Hairong Tian<sup>3†</sup>, Shuting Xie<sup>1</sup>, Ye Ji<sup>3</sup>, Ziyun Li<sup>3</sup>, Junxi Lu<sup>1</sup>, Huijuan Lu<sup>1</sup>, Bo Liu<sup>3</sup> and Fang Liu<sup>1,4\*</sup> 

## Abstract

**Background:** Complement factor H (CFH) has been found to be associated with insulin resistance. This study assessed the correlation between CFH and other clinical parameters, and determined whether CFH played a role in gestational diabetes mellitus (GDM) and adverse pregnancy outcomes.

**Methods:** A total of 397 pregnant women were included for analysis in this nested case-control study. Clinical parameters and serum were collected within the 11–17th gestational age at the first prenatal visit. At 24–28 weeks of gestation, a 75 g oral glucose tolerance test was performed and subjects were divided into a GDM ( $n = 80$ ) and a non-GDM control group ( $n = 317$ ). The delivery data were also followed. The serum CFH level was assayed by ELISA.

**Results:** CFH was higher in GDM than in non-GDM controls (280.02 [58.60] vs. 264.20 [68.77];  $P = 0.014$ ). CFH level was moderately associated with pre-pregnancy body mass index (BMI), BMI and total triglycerides (TG), and slightly associated with gestational age, low density lipoprotein cholesterol (LDL-C), total cholesterol (TC) in GDM and non-GDM (all  $P < 0.05$ ). Moreover, CFH level was moderately correlated with alkaline phosphatase (ALP) and slightly correlated with age, uric acid (UA) and total bilirubin (TB) in non-GDM (all  $P < 0.05$ ). After adjustment for clinical confounding factors, BMI, TG, gestational age, ALP, TB, age and UA were independent risk factors for  $\log_{10}$  CFH levels (all  $P < 0.05$ ) in all subjects. In addition, overweight or obese pregnant women, women with hypertriglyceridemia and women in the second trimester had significantly higher CFH levels than normal weight and underweight group ( $P < 0.001$ ), the non-hypertriglyceridemia group ( $P < 0.001$ ) and women in the first trimester group ( $P < 0.05$ ) in all pregnant women respectively. Following binary logistic regression, CFH was not independently associated with GDM and related pregnant outcomes.

**Conclusions:** The CFH in 11–17th weeks of gestation might be affected by many factors, including BMI, TG, gestational age, ALP, TB, age and UA. CFH was not an independent risk factor for GDM and adverse pregnancy outcomes.

**Keywords:** Complement factor H, Gestational diabetes mellitus, Pregnancy

\* Correspondence: [f-liu@sjtu.edu.cn](mailto:f-liu@sjtu.edu.cn)

<sup>†</sup>Junxian Li, Ying Shen and Hairong Tian contributed equally to this work.

<sup>1</sup>Department of Endocrinology & Metabolism, Shanghai Jiao-Tong University Affiliated Sixth People's Hospital, Shanghai Key Laboratory of Diabetes, Shanghai Clinical Medical Center of Diabetes, Shanghai Key Clinical Center of Metabolic Diseases, Shanghai Institute for Diabetes, Shanghai 200233, China

<sup>4</sup>Department of Endocrinology and Metabolism, Shanghai General Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai 200080, China

Full list of author information is available at the end of the article



© The Author(s). 2021 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

## Background

Gestational diabetes mellitus (GDM) is a condition in which glucose intolerance appears anytime during pregnancy leading to rise in blood glucose levels. The incidence of GDM varies according to diagnostic criteria, geographic regions and race/ethnicity. In addition, its incidence is expected to increase in the future [1–3].

The pathogenesis of GDM includes insulin resistance and insufficient insulin secretion, but the specific mechanism remains unclear [3]. Gestational hyperglycemia has serious adverse consequences on pregnant mothers, the developing fetus and neonates, including cesarean section, macrosomia, and premature rupture of membranes (PROM). Long-term consequences from GDM include development of cardiovascular disease (CVD) and type 2 diabetes mellitus (T2DM) in both mothers and offsprings and additionally obesity in the offsprings [3, 4]. Therefore, a coordinated study on the disease pathogenesis of GDM and predictive biomarkers is of great significance.

Previous researches found that inflammation might play a key role in the pathogenesis of GDM and inflammatory cytokines were predictive biomarkers of GDM [5]. For instance, Ueland et al. found that the macrophage marker sCD163 increased at 14–16th weeks of gestation [6], and Ozgu-Erdinc et al. reported that C-reactive protein (CRP) increased within 11–14th weeks of gestation [7], and both inflammatory markers were independently associated with GDM. On the contrary, other researchers found that combination of clinical factors and biomarkers such as TNF- $\alpha$  and high sensitivity-CRP did not show significant improvement in the prediction of GDM [8, 9].

The complement system is an important part of innate immunity, and its activation occurs through three distinct pathways: the classical pathway, the lectin pathway and the alternative pathway [10]. Human complement factor H (CFH) is a soluble complement system inhibitor and can protect cells and tissues from unexpected complement system-mediated damage [11]. The gene that encodes CFH is located on chromosome 1q31.3 and is mainly expressed by the liver [12, 13], and other cell types including endothelial cells [14], retinal pigment epithelial cells [15], and adipocytes [16, 17]. CFH levels in the plasma varied widely from 116 to 562  $\mu\text{g/ml}$  depending on genetic and environmental factors [11, 18], and might even increase in pregnant women [19].

It has been suggested that CFH was associated with obesity and metabolic disorders. Moreno-Navarrete et al. found that the CFH level significantly increased in patients with altered glucose tolerance and T2DM, and plasma CFH levels were negatively associated with insulin sensitivity [16, 20]. It was considered that attenuated insulin sensitivity represents the main

pathogenic mechanism in GDM, and thus CFH might be related to GDM development.

Recently, Shen et al. revealed that complement system-associated proteins, including CFH, changed significantly in GDM at 12–14th gestational age as measured by proteomic analysis [21]. Therefore, the role of CFH in GDM patients requires further study.

Thus, the aim of this study was to assess the correlation between CFH and other clinical parameters in Chinese pregnant women, and to determine whether CFH played a role in GDM and adverse pregnancy outcomes.

## Methods

### Study population

It was a nested case-control study. Pregnant women were recruited in a prospective cohort and drawn blood samples at the first prenatal visit (< 24th gestational age). Inclusion criteria included the following: the first prenatal visit that was less than 24 weeks gestation; do not smoke or consume alcohol; no pre-existing medical disorders including diabetes and acute or chronic inflammation. A total of 607 women who met the inclusion criteria were recruited at the first prenatal visit. At 24–28th weeks of gestation, all women experienced the 75-g oral glucose tolerance test (75-g OGTT) and GDM was diagnosed if one of the following criteria was met or exceeded: 0 h glucose  $\geq 5.1$  mmol/L, 1 h glucose  $\geq 10$  mmol/L, and 2 h glucose  $\geq 8.5$  mmol/L [22]. Clinical and biochemical data from the first prenatal visit to delivery were collected at the Department of Obstetrics and Gynecology and the Department of Endocrinology and Metabolism of the Jin Shan Branch of Shanghai Sixth People's Hospital, from February 2017 to April 2019. Subsequently, a total of 210 were excluded due to pre-conception diabetes ( $n = 4$ ), twin pregnancy ( $n = 4$ ) and incomplete clinical or measurement data ( $n = 202$ ). The final number of women included for analysis was 397, and those women's first prenatal visits were within the 11–17th gestational age.

### Data and serum sample collection

All pregnant women who met the inclusion criteria completed questionnaires (Additional file 1) that collected general background information including age, last menstrual period, reproductive history, and family history of diabetes at the first prenatal visit. Moreover, height, weight, and systolic and diastolic blood pressure were recorded on a standardized form by the physician during the examination. Pre-pregnancy body mass index (BMI) was calculated as pre-pregnancy body weight (in kg)/height<sup>2</sup> (in m<sup>2</sup>). BMI was calculated at point of first prenatal visit and was calculated as body weight (in kg)/height<sup>2</sup> (in m<sup>2</sup>). Each participant was drawn 3 ml venous

blood following one night of fasting at the first prenatal visit, and serum samples were obtained aseptically by centrifugation at 3500 rpm for 15 min, which were then frozen at  $-80^{\circ}\text{C}$  until being used [23]. Macrosomia was defined as birth weight  $\geq 4000$  g. Premature rupture of membrane (PROM) was defined as rupture of membranes before the onset of labour [24]. Estimated blood loss at delivery was defined as volume of blood loss from women during delivery within first 24 h after birth, and was calculated by the following ways: gauzes and pads with blood were weighed and an equivalent volume was estimated; blood volume in the suction bottle was measured.

#### Laboratory measurements

Plasma glucose values were measured by the glucose oxidase method. HbA1c was determined by high-pressure liquid chromatography. Glycated serum albumin (GA) was tested by the liquid enzymatic assay. Other biochemical indices evaluating hepatic and renal functions such as aspartate aminotransferase (AST), alanine aminotransferase (ALT), total bilirubin (TB),  $\gamma$ -glutamyltransferase ( $\gamma$ -GT), alkaline phosphatase (ALP), creatinine (Cr), blood urea nitrogen (BUN), and uric acid (UA) were performed by enzymatic methods. Serum lipids including total triglycerides (TG), total cholesterol (TC) and low density lipoprotein cholesterol (LDL-C) were also tested by enzymatic methods. Serum albumin (ALB) was measured by the Bromocresol Green (BCG) dye-binding method. Then serum CFH concentrations were measured in duplicate by using MicroVue Factor H EIA kits (Quidel Corporation, USA) according to the manufacturer's instructions, and the detectable quantitation range was 4.64–521 ng/ml. The intra- and inter-assay coefficients of variation (CV) were less than 10%. The control values were within the control ranges.

#### Statistical analysis

Data are expressed as median (interquartile range, [IQR]) for continuous variables with non-normal distribution, mean  $\pm$  standard deviation (SD) for continuous variables with normal distribution, and percentage (%) for categorical variables. Differences between groups were evaluated with the Chi-square test for categorical variables and the Student's t-test or Mann-Whitney U test for continuous variables. The correlation between CFH and other characteristics at the first prenatal visit was evaluated with the Spearman's rank correlation and multiple stepwise linear regression analysis. Binary logistic regression analysis was performed to evaluate the odds ratios (OR) and 95% confidence intervals (CIs) in multivariable analysis. All statistical analyses were measured by SPSS version 26.0 (SPSS Inc., Chicago, IL,

USA). A two-sided alpha value of  $P < 0.05$  was considered statistically significant.

## Results

### Subject characteristics

In this nested case-control study, 397 women completed the study and were assigned into a GDM ( $n = 80$ ) and a non-GDM group ( $n = 317$ ) based on the 75 g OGTT results at 24–28th gestational age. The clinical and biochemical characteristics of both are shown in Table 1. There were significant differences in terms of age, pre-pregnancy BMI, BMI, FPG, HbA1c, ALT, UA, TG, TC, LDL-C and CFH (all at  $P < 0.05$ ) between the GDM and non-GDM controls. After comparison of pregnant outcomes, the incidence of macrosomia (%) was significantly higher in GDM than in non-GDM controls ( $P < 0.05$ ). However the incidence of caesarean section (%), PROM (%), fetal distress (%), and other outcomes such as gestational age at delivery, estimated blood loss at delivery, and the Apgar score showed no differences between GDM and non-GDM controls.

### The association between CFH and other clinical and biochemical characteristics

To assess the relationship between CFH and other parameters, the Spearman's correlation analysis was used to derive a correlation coefficient ( $r$ ). The result showed that CFH was found to be significantly moderately positively ( $0.3 \leq r < 0.5$ ) associated with pre-pregnancy BMI, BMI and TG, and significantly slightly positively ( $r < 0.3$ ) associated with gestational age, LDL-C, TC in GDM, non-GDM (Table 2). In addition, this study showed that the CFH level was moderately correlated with ALP and slightly correlated with age, UA and TB (all  $P < 0.05$ ) in non-GDM (Table 2).

The pregnant women were divided into three categories by BMI: an underweight category ( $\text{BMI} < 18.5 \text{ kg/m}^2$ ), a normal weight category ( $18.5 < \text{BMI} < 23.9 \text{ kg/m}^2$ ), and an overweight or obese category ( $\text{BMI} \geq 24 \text{ kg/m}^2$ ). The result showed that overweight or obese pregnant women had significantly higher levels of CFH as compared to normal and underweight pregnant women in GDM (303.45 [58.39] vs. 270.29 [61.63] vs. 259.00 [54.13], respectively,  $P < 0.01$ ), in non-GDM controls (294.93 [68.32] vs. 263.13 [58.63] vs. 229.55 [49.70], respectively,  $P < 0.001$ ) and in all pregnant women (296.49 [65.11] vs. 263.43 [59.61] vs. 230.93 [51.36], respectively,  $P < 0.001$ ) (Fig. 1). Hypertriglyceridemia is a lipid metabolism disorder, so the subjects were divided into two categories by TG: a non-hypertriglyceridemia category ( $\text{TG} < 1.7 \text{ mmol/L}$ ) and a hypertriglyceridemia category ( $\text{TG} \geq 1.7 \text{ mmol/L}$ ). Participants with hypertriglyceridemia had significantly higher CFH levels than non-hypertriglyceridemia participants in non-GDM (290.23 [85.29] vs. 259.58 [59.26], respectively,

**Table 1** Comparison of the clinical characteristics of pregnant women with and without gestational diabetes mellitus (GDM)

Parameters <sup>a</sup>	GDM (n = 80)	Non-GDM (n = 317)	P value <sup>b</sup>
	Median (IQR) or mean ± SD or %	Median (IQR) or mean ± SD or %	
Gestational age at the first prenatal visit, week	14.00 (2.75)	13.00 (3.00)	0.292 <sup>†</sup>
Age, years	29.00 (5.00)	27.00 (5.00)	0.012 <sup>†</sup>
Pre-pregnancy BMI, kg/m <sup>2</sup>	21.70 (4.33)	20.80 (3.65)	< 0.001 <sup>†</sup>
BMI, kg/m <sup>2</sup>	21.98 (4.93)	20.90 (3.75)	< 0.001 <sup>†</sup>
SBP, mmHg	118.00 (16.75)	115.00 (14.00)	0.215 <sup>†</sup>
DBP, mmHg	75.00 (11.00)	74.00 (12.50)	0.502 <sup>†</sup>
Family History of diabetes, %	10	4.7	0.125 <sup>#</sup>
GA, %	12.08 ± 1.91	11.92 ± 1.55	0.422 <sup>*</sup>
FPG, mmol/L	4.85 (0.60)	4.70 (0.50)	< 0.001 <sup>†</sup>
HbA1c, %	5.20 (0.40)	5.10 (0.30)	< 0.001 <sup>†</sup>
ALT, units/L	14.50 (16.50)	12.00 (11.00)	0.024 <sup>†</sup>
AST, units/L	16.00 (7.60)	16.00 (5.30)	0.940 <sup>†</sup>
γ-GT, units/L	11.50 (8.30)	11.00 (8.30)	0.425 <sup>†</sup>
ALP, units/L	45.00 (15.75)	46.00 (14.00)	0.488 <sup>†</sup>
TB, μmol/L	8.20 (4.58)	7.90 (3.75)	0.539 <sup>†</sup>
ALB, g/L	42.10 (3.05)	42.100 (3.65)	0.872 <sup>†</sup>
BUN, mmol/L	2.80 (0.90)	2.70 (0.80)	0.458 <sup>†</sup>
Cr, μmol/L	44.00 (8.00)	44.00 (8.00)	0.197 <sup>†</sup>
UA, μmol/L	220.50 (62.00)	204.00 (60.00)	0.004 <sup>†</sup>
TG, mmol/L	1.49 (0.79)	1.295 (0.65)	0.007 <sup>†</sup>
TC, mmol/L	4.62 ± 0.79	4.42 ± 0.78	0.042 <sup>*</sup>
LDL-C, mmol/L	2.52 ± 0.67	2.36 ± 0.66	0.051 <sup>*</sup>
CFH, μg/ml	280.02 (58.60)	264.20 (68.77)	0.014 <sup>†</sup>
GDM screening 75 g OGTT			
Gestational age at 75 g OGTT, week	25.00 (2.00)	25.00 (1.00)	0.777 <sup>†</sup>
Glucose 0 h, mmol/L	5.15 (0.70)	4.50 (0.50)	< 0.001 <sup>†</sup>
Glucose 1 h, mmol/L	9.79 ± 1.82	7.24 ± 1.39	< 0.001 <sup>*</sup>
Glucose 2 h, mmol/L	7.95 (2.15)	6.31 (1.56)	< 0.001 <sup>†</sup>
Pregnancy outcomes			
Gestational age at delivery, week	39.00 (1.00)	39.00 (1.00)	0.100 <sup>†</sup>
Caesarean section, %	48.8	42.3	0.296 <sup>#</sup>
PROM, %	11.3	11.4	0.979 <sup>#</sup>
Estimated blood loss at delivery, ml	300.00 (90.00)	300.00 (100.00)	0.71 <sup>†</sup>
Macrosomia, %	11.0	4.4	0.038 <sup>#</sup>
Apgar score	10.00 (0.00)	10.00 (0.00)	0.291 <sup>†</sup>

**Abbreviations:** GDM gestational diabetes mellitus; BMI body mass index; SBP systolic blood pressure; DBP diastolic blood pressure; GA Glycated serum albumin; FPG fasting plasma glucose; HbA1c glycosylated hemoglobin A1c; ALT alanine aminotransferase; AST aspartate aminotransferase; γ-GT γ-glutamyltransferase; ALP alkaline phosphatase; TB total bilirubin; ALB albumin; BUN blood urea nitrogen; Cr creatinine; UA uric acid; TC total cholesterol; TG total triglycerides; LDL-C low-density lipoprotein cholesterol; CFH complement factor H; PROM premature rupture of membrane

a. Data are expressed as median (interquartile range, [IQR]) for continuous variables with non-normal distribution, mean ± standard deviation (SD) for continuous variables with normal distribution, and percentage (%) for categorical variables

b. <sup>†</sup>Derived from Student's t-test. <sup>\*</sup>Derived from Mann-Whitney U test. <sup>#</sup>Derived from Chi-square test

$P < 0.001$ ) and in all pregnant women (291.55 [75.46] vs. 260.60 [60.28], respectively,  $P < 0.001$ ) (Fig. 1). In addition, the CFH level was also significantly higher in the second trimester (13 ~ 28 gestational age) than in the first

trimester (0 ~ 12 gestational age) in non-GDM (265.25 [64.09] vs. 252.19 [71.24], respectively,  $P = 0.037$ ) and in all pregnant women (268.04 [60.96] vs. 256.81 [71.68], respectively,  $P = 0.019$ ) (Fig. 1).

**Table 2** Correlation between complement factor H (CFH) and other variables in the first prenatal visit

	CFH ( $\mu\text{g/ml}$ )			
	GDM ( $n = 80$ )		Non-GDM ( $n = 317$ )	
	r	P	r	P
Gestational age at the first prenatal visit, week	0.235	0.036	0.141	0.012
Age, years	0.076	0.504	0.199	< 0.001
Pre-pregnancy BMI, $\text{kg/m}^2$	0.319	0.004	0.344	< 0.001
BMI, $\text{kg/m}^2$	0.355	0.001	0.366	0.000
SBP, mmHg	0.214	0.057	0.104	0.064
DBP, mmHg	0.167	0.140	0.099	0.079
Family history of diabetes, %	0.000	1.000	0.164	0.003
GA, %	0.072	0.524	-0.083	0.139
FPG, mmol/L	0.003	0.979	0.095	0.091
HbA1c, %	0.191	0.089	0.174	0.002
ALT, units/L	0.083	0.464	0.035	0.530
AST, units/L	0.103	0.365	0.018	0.750
$\gamma$ -GT, units/L	0.070	0.537	0.109	0.052
ALP, units/L	0.127	0.263	0.392	< 0.001
TB, $\mu\text{mol/L}$	-0.142	0.208	-0.218	< 0.001
ALB, g/L	-0.037	0.747	-0.081	0.148
BUN, mmol/L	0.005	0.966	-0.093	0.101
Cr, $\mu\text{mol/L}$	-0.078	0.491	0.017	0.766
UA, $\mu\text{mol/L}$	0.142	0.209	0.231	< 0.001
TG, mmol/L	0.312	0.005	0.319	< 0.001
TC, mmol/L	0.235	0.038	0.189	0.001
LDL-C, mmol/L	0.241	0.034	0.189	0.001

**Abbreviations:** CFH complement factor H; GDM gestational diabetes mellitus; r correlation coefficient; BMI body mass index; SBP systolic blood pressure; DBP diastolic blood pressure; GA Glycated serum albumin; FPG fasting plasma glucose; HbA1c glycosylated hemoglobin A1c; ALT alanine aminotransferase; AST aspartate aminotransferase;  $\gamma$ -GT  $\gamma$ -glutamyltransferase; ALP alkaline phosphatase; TB total bilirubin; ALB albumin; BUN blood urea nitrogen; Cr creatinine; UA uric acid; TG total triglycerides; TC total cholesterol; LDL-C low-density lipoprotein cholesterol; TB total bilirubin. Data were derived from Spearman's rank correlation analysis

### Multiple stepwise linear regression analysis of possible independent risk factors of $\log_{10}$ CFH

To determine which factors were independently associated with serum CFH levels, the multiple stepwise linear regression was performed in all subjects. Clinical parameters including gestational age, age, BMI, SBP, DBP, family history of diabetes, FPG, ALT, AST,  $\gamma$ -GT, ALP, TB, ALB, BUN, UA, TG, TC and LDL-C were included in analysis of multiple stepwise linear regression. This analysis revealed that BMI (Standardized Coefficients Beta [ $\beta$ ] = 0.230,  $P < 0.001$ ), TG ( $\beta = 0.130$ ,  $P = 0.011$ ), gestational age ( $\beta = 0.138$ ,  $P = 0.004$ ), ALP ( $\beta = 0.197$ ,  $P < 0.001$ ), TB ( $\beta = -0.174$ ,  $P < 0.001$ ), age ( $\beta = 0.111$ ,  $P = 0.020$ ), and UA ( $\beta = 0.106$ ,  $P = 0.027$ ) were independent risk factors for serum  $\log_{10}$  CFH levels in all pregnant women.

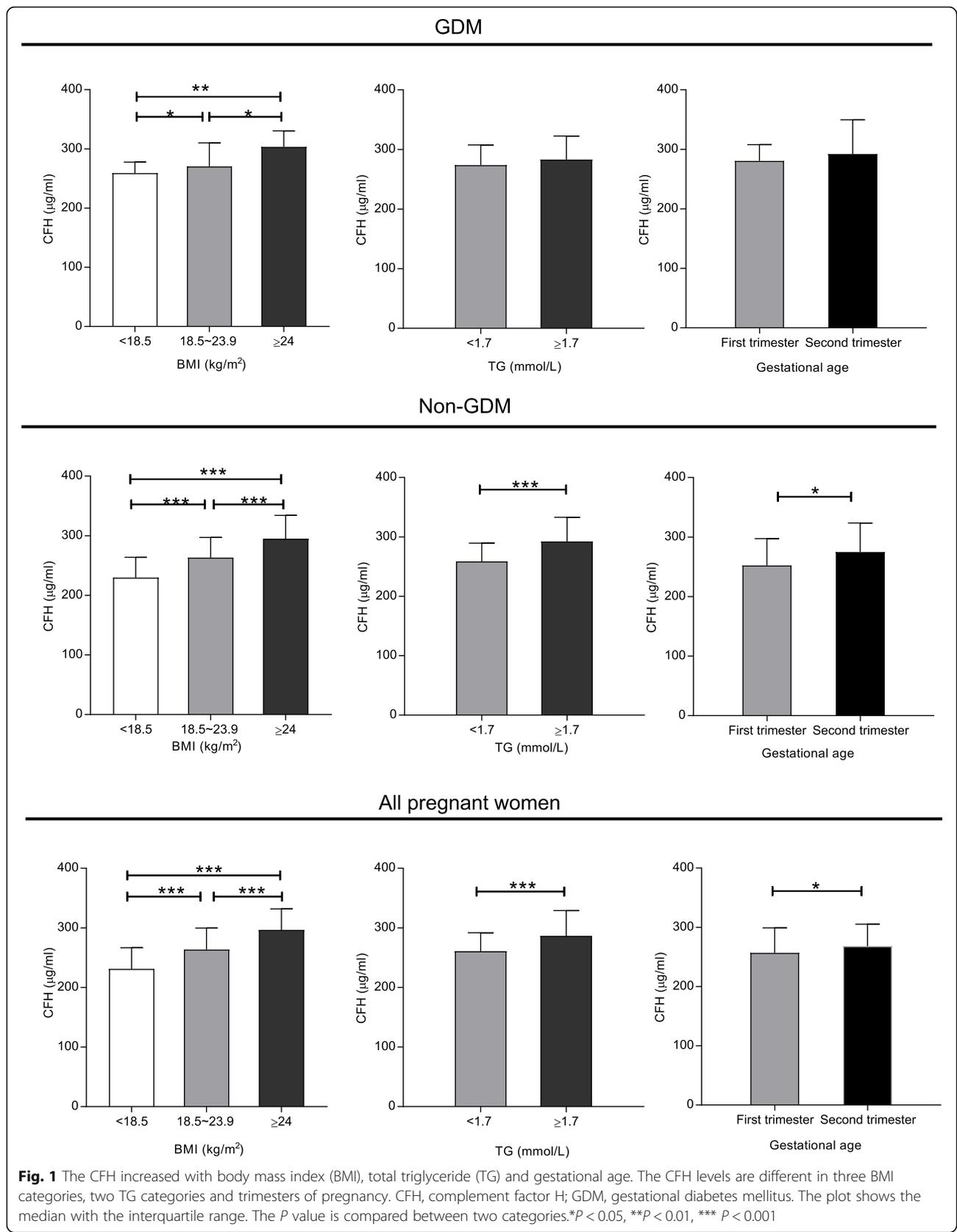
### Binary logistic regression analysis of factors affecting GDM and adverse pregnancy outcomes

To determine whether CFH was independently associated with GDM and undesirable pregnancy outcomes or

not, binary logistic regression was performed (Table 3). However, no significantly independent association was found between serum CFH and GDM and adverse pregnancy outcomes. In addition, BMI was independently associated with GDM (OR, 1.090; 95% CI, 1.005–1.183;  $P = 0.037$ ). In the macrosomia subgroup, BMI was also an independent risk factor for macrosomia development (OR, 1.203; 95% CI, 1.065–1.360;  $P = 0.003$ ). In the caesarean subgroup, age (OR, 1.127; 95% CI, 1.068–1.190;  $P < 0.001$ ), BMI (OR, 1.092; 95% CI, 1.011–1.179;  $P = 0.026$ ) and ALT (OR, 1.024; 95% CI, 1.006–1.043;  $P = 0.008$ ) were independent risk factors. We did not identify any factor that was significantly correlated with PROM in this study.

### Discussion

The CFH was found to be higher in GDM as compared with non-GDM controls in Chinese women. In addition, the CFH was independently associated with BMI, TG, gestational age, ALP, age, TB and UA in all subjects.



**Table 3** Binary logistic regression analysis of factors affecting gestational diabetes mellitus (GDM) and adverse pregnancy outcomes

	<b>GDM</b> <b>OR (95% CI)</b>	<b>Macrosomia</b> <b>OR (95% CI)</b>	<b>Caesarean section</b> <b>OR (95% CI)</b>	<b>PROM</b> <b>OR (95% CI)</b>
CFH	1.002 (0.996–1.007)	0.993 (0.982–1.003)	0.997 (0.992–1.002)	0.997 (0.990–1.005)
Age	1.042 (0.981–1.107)	0.966 (0.868–1.074)	1.127 (1.068–1.190)***	1.033 (0.959–1.113)
BMI	1.090 (1.005–1.183)*	1.203 (1.065–1.360)**	1.092 (1.011–1.179)*	0.994 (0.890–1.110)
Family history of diabetes	1.457 (0.551–3.853)	0.385 (0.046–3.233)	2.041 (0.756–5.510)	0.708 (0.150–3.336)
ALT	1.008 (0.991–1.026)	1.019 (0.993–1.045)	1.024 (1.006–1.043)**	1.002 (0.979–1.026)
ALP	0.995 (0.971–1.020)	1.008 (0.968–1.050)	0.995 (0.974–1.017)	1.009 (0.978–1.041)
TB	1.035 (0.946–1.133)	0.842 (0.706–1.004)	1.045 (0.968–1.129)	0.978 (0.871–1.099)
UA	1.003 (0.997–1.009)	1.000 (0.990–1.009)	1.002 (0.997–1.007)	1.000 (0.993–1.008)
TG	1.211 (0.783–1.874)	1.312 (0.647–2.660)	1.021 (0.684–1.523)	0.705 (0.373–1.332)
TC	1.123 (0.789–1.599)	0.980 (0.545–1.764)	1.074 (0.794–1.451)	1.429 (0.926–2.206)

*Abbreviations:* GDM gestational diabetes mellitus; PROM premature rupture of the membrane; OR odds ratio; 95% CI 95% confidence interval; CFH complement factor H; BMI body mass index; ALT alanine aminotransferase; ALP alkaline phosphatase; TB total bilirubin; UA uric acid; TG total triglycerides; TC total cholesterol. Data were derived from binary logistic regression. \* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$

Following binary logistic regression, CFH was not independently associated with GDM and pregnancy outcomes. There are some strengths in this study. First, the research tested CFH firstly in pregnant women whose prenatal and delivery clinical data were also followed. Second, this study found some clinical parameters that might independently affect CFH levels and the complement system activity during pregnancy.

Some results of this study were generally consistent with previous work. This study and that of others found that CFH was positive associated with BMI [16, 25, 26], and fasting TG [16]. It was reported [16, 25] that alternative complement activation was associated with elevated BMI and the synthesis of TG in adipocytes, because C3, C3a, and other alternative pathway components were all associated with BMI [25, 26], and the C3a degradation product C3a des-Arg could stimulate the synthesis of TG in adipocytes further [27]. Since CFH is a complement alternative pathway inhibitor, it might represent a compensatory increase when the complement alternative pathway system is activated, which could lead to CFH being positively correlated with BMI and TG levels.

According to the criteria for defining obesity and hypertriglyceridemia in China [28, 29], the subjects were divided into three categories by BMI, and two categories by TG, as described in the results section above. Our results indicated that overweight or obese pregnant women, and women with hypertriglyceridemia, had significantly higher levels of CFH as compared with other categories, which provides a novel conceptual framework for determining the impact of overweight, obesity and hypertriglyceridemia on regulating the complement system.

Previous researches have reported that CFH elevated in pregnancy [19], and this study found that CFH level

increased with the gestational age. Since CFH is a complement system inhibitor, it might be a mechanism of immunosuppression in pregnancy.

Moreover, we found that the CFH level had a moderately positive association with ALP, and ALP was an independent risk factor for serum CFH levels. As we know, ALP in pregnant mothers is mainly derived from placental tissues, the liver and bone [30, 31], and these tissues also affect the complement system and CFH expression [32–35]. This connection might account for the positive association between serum CFH and ALP.

This study also demonstrated that CFH was slightly ( $r < 0.3$ ) associated with TB, age, and UA, and these factors might be independent risk factors for CFH levels in all subjects. The CFH level was slightly negatively associated with TB. Basiglio et al. reviewed that unconjugated bilirubin could inhibit activation of the complement system by preventing complement factor C1q interacting with immunoglobulins, and this might decrease CFH levels when the complement system was inhibited [36–38]. CFH was slightly positively correlated with age, which could be attributed to normal physiological phenomenon since previously published literature reported that the CFH level was significantly higher in adults than in neonates [39]. Previous work also similarly showed that UA was positively connected with complement C3 in adults, and that UA could stimulate the expression of complement C3 in a dose-dependent fashion [40]. Thus, the rising CFH levels might be a compensatory reaction after UA stimulated the complement system.

Some results of this study were not generally consistent with previous work conducted in Chinese females. Shen et al. used proteomic analysis and found that CFH changed significantly in GDM as compared with non-GDM controls at 12–16th gestational age after adjusting for maternal age, gravity, parity, BMI, gestational age at

delivery and gestational age at time of sample collection [21]. However, our study found that there was no significant difference of CFH levels on comparing GDM and non-GDM after adjusting for other clinical characteristics. This discordance could be caused by the fact that the case numbers of the GDM group were relatively small and the detection methods of CFH were different.

In the Moreno-Navarrete et al. study, the CFH level was negatively associated with insulin sensitivity [16], therefore GDM patients with insulin resistance were speculated to have elevated CFH levels. Although our study found the CFH level was significantly higher in GDM than non-GDM controls, CFH was not independently related to GDM and adverse pregnancy outcomes. Therefore, it is rational to consider that although CFH is positive related with insulin resistance, it is not independent risk factors of insulin resistance. Insulin resistance is commonly exhibited in GDM, impaired glucose tolerance and T2DM, and these conditions are more likely to have high BMI and TG. In other words, it might be possible that the body adipose component and TG, but not the resulting CFH alterations, independently and directly influence insulin resistance in pregnancy.

This study had some limitations. First, the current study recruited a relatively small sample size of women with progressive GDM. Second, the lack of data reflecting islet  $\beta$  cell function such as fasting insulin and C-peptide levels resulted in the defect of the putative association between CFH and insulin resistance during pregnancy.

## Conclusion

This study helps advance our understanding of the role of CFH and the complement system in GDM and pregnancy. The data showed that the CFH level was positively associated with BMI, TG, gestational age, ALP, age and UA, and was negatively correlated with TB. These factors were independent risk factors for CFH levels which might affect the complement system activity when women are pregnant. However, CFH levels were not independently correlated with GDM and adverse pregnancy outcomes. Future studies of the associations between CFH and insulin resistance in pregnancy are indeed warranted.

## Abbreviations

CFH: Complement factor H; GDM: gestational diabetes mellitus; TG: total triglycerides; BMI: body mass index; PROM: premature rupture of membranes; CVD: cardiovascular disease; T2DM: type 2 diabetes mellitus; CRP: C-reactive protein; 75-g OGTT: 75-g oral glucose tolerance test; GA: Glycated serum albumin; AST: aspartate aminotransferase; ALT: alanine aminotransferase; TB: total bilirubin;  $\gamma$ -GT:  $\gamma$ -glutamyltransferase; ALP: alkaline phosphatase; Cr: creatinine; BUN: blood urea nitrogen; UA: uric acid; TC: total cholesterol; LDL-C: low density lipoprotein cholesterol; ALB: albumin; BCG: Bromocresol Green; CV: coefficients of variation; OR: odds ratios; CIs: confidence intervals; SBP: systolic blood pressure; DBP: diastolic blood pressure; FPG: fasting plasma glucose; HbA1c: glycosylated hemoglobin A1c; IQR: Interquartile range; SD: standard deviation

## Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12884-021-04031-w>.

### Additional file 1.

## Acknowledgments

We thank all other investigators, study staff and all participants of the present study for their valuable contributions.

## Authors' contributions

FL designed the study. HRT, JXL1, YS and STX contributed to sample collection and data analysis. JXL1 drafted the manuscript. FL reviewed and edited the manuscript. JXL2 and HJL measured biochemical indices. HRT, JXL1, YS, STX, YJ, ZYL, and BL collected all samples and the clinical data, and took responsibility of data integrity. All authors reviewed the manuscript. All authors read and approved the final manuscript.

## Funding

This work was supported by the National Natural Science Foundation of China (Grant No. 81770802 for Fang Liu). The funding source had no role in the design of the study, collection, analysis, and interpretation of data or in writing the manuscript.

## Availability of data and materials

The datasets generated and analyzed during the current reported work are available from the corresponding authors upon reasonable request.

## Declarations

### Ethics approval and consent to participate

The study was approved by the local Ethics Committee of Jin Shan Branch of Shanghai Sixth People's Hospital and performed in accordance with the ethical standards that were laid down in the 1964 Declaration of Helsinki and subsequent amendments. Written informed consent was obtained from each participant in this study.

### Consent for publication

Not applicable.

### Competing interests

The authors declare that they have no competing interests.

### Author details

<sup>1</sup>Department of Endocrinology & Metabolism, Shanghai Jiao-Tong University Affiliated Sixth People's Hospital, Shanghai Key Laboratory of Diabetes, Shanghai Clinical Medical Center of Diabetes, Shanghai Key Clinical Center of Metabolic Diseases, Shanghai Institute for Diabetes, Shanghai 200233, China. <sup>2</sup>Department of Endocrinology & Metabolism, The Affiliated Jiangsu Shengze Hospital of Nanjing Medical University, Suzhou 215228, China. <sup>3</sup>Department of Endocrinology and Metabolism, Jin Shan Branch of Shanghai Sixth People's Hospital, Shanghai 201599, China. <sup>4</sup>Department of Endocrinology and Metabolism, Shanghai General Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai 200080, China.

Received: 16 December 2020 Accepted: 2 August 2021

Published online: 17 August 2021

## References

- Lee KW, Ching SM, Ramachandran V, Yee A, Hoo FK, Chia YC, et al. Prevalence and risk factors of gestational diabetes mellitus in Asia: a systematic review and meta-analysis. *BMC Pregnancy Childbirth*. 2018; 18(1):494.
- Lean SC, Derricott H, Jones RL, Heazell AEP. Advanced maternal age and adverse pregnancy outcomes: a systematic review and meta-analysis. *PLoS One*. 2017;12(10):e0186287.
- Plows JF, Stanley JL, Baker PN, Reynolds CM, Vickers MH. The Pathophysiology of Gestational Diabetes Mellitus. *Int J Mol Sci*. 2018;19(11):3342.

4. Metzger BE, Lowe LP, Dyer AR, Trimble ER, Chaovarindr U, Coustan DR, et al. Hyperglycemia and adverse pregnancy outcomes. *N Engl J Med*. 2008; 358(19):1991–2002.
5. Lekva T, Norwitz ER, Aukrust P, Ueland T. Impact of systemic inflammation on the progression of gestational diabetes mellitus. *Curr Diab Rep*. 2016; 16(4):26.
6. Ueland T, Michelsen AE, Aukrust P, Henriksen T, Bollerslev J, Lekva T. Adipokines and macrophage markers during pregnancy-possible role for sCD163 in prediction and progression of gestational diabetes mellitus. *Diabetes Metab Res Rev*. 2019;35(3):e3114.
7. Ozgu-Erdinc AS, Yilmaz S, Yeral MI, Seckin KD, Erkaya S, Danisman AN. Prediction of gestational diabetes mellitus in the first trimester: comparison of C-reactive protein, fasting plasma glucose, insulin and insulin sensitivity indices. *J Matern Fetal Neonatal Med*. 2015;28(16):1957–62.
8. Syngelaki A, Visser GH, Krithinakis K, Wright A, Nicolaides KH. First trimester screening for gestational diabetes mellitus by maternal factors and markers of inflammation. *Metabolism*. 2016;65(3):131–7.
9. Powe CE. Early pregnancy biochemical predictors of gestational diabetes mellitus. *Curr Diab Rep*. 2017;17(2):12.
10. Ricklin D, Reis ES, Lambris JD. Complement in disease: a defence system turning offensive. *Nat Rev Nephrol*. 2016;12(7):383–401.
11. Ferreira VP, Pangburn MK, Cortés C. Complement control protein factor H: the good, the bad, and the inadequate. *Mol Immunol*. 2010;47(13):2187–97.
12. Pouw RB, Vredevoogd DW, Kuijpers TW, Wouters D. Of mice and men: the factor H protein family and complement regulation. *Mol Immunol*. 2015; 67(1):12–20.
13. Schwaebler W, Zwirner J, Schulz TF, Linke RP, Dierich MP, Weiss EH. Human complement factor H: expression of an additional truncated gene product of 43 kDa in human liver. *Eur J Immunol*. 1987;17(10):1485–9.
14. Broomans RA, van der Ark AA, Buurman WA, van Es LA, Daha MR. Differential regulation of complement factor H and C3 production in human umbilical vein endothelial cells by IFN-gamma and IL-1. *J Immunol*. 1990;144(10):3835–40.
15. Chen M, Forrester JV, Xu H. Synthesis of complement factor H by retinal pigment epithelial cells is down-regulated by oxidized photoreceptor outer segments. *Exp Eye Res*. 2007;84(4):635–45.
16. Moreno-Navarrete JM, Martínez-Barricarte R, Catalán V, Sabater M, Gómez-Ambrosi J, Ortega FJ, et al. Complement factor H is expressed in adipose tissue in association with insulin resistance. *Diabetes*. 2010;59(1):200–9.
17. Lehr S, Hartwig S, Lamers D, Famulla S, Müller S, Hanisch FG, et al. Identification and validation of novel adipokines released from primary human adipocytes. *Mol Cell Proteomics*. 2012;11(11):M111.010504.
18. Esparza-Gordillo J, Soria JM, Buil A, Almasy L, Blangero J, Fontcuberta J, et al. Genetic and environmental factors influencing the human factor H plasma levels. *Immunogenetics*. 2004;56(2):77–82.
19. Bohács A, Bikov A, Ivancsó I, Czaller I, Böcskei R, Müller V, et al. Relationship of circulating C5a and complement factor H levels with disease control in pregnant women with asthma. *Respir Care*. 2016;61(4):502–9.
20. Moreno-Navarrete JM, Fernández-Real JM. The complement system is dysfunctional in metabolic disease: evidences in plasma and adipose tissue from obese and insulin resistant subjects. *Semin Cell Dev Biol*. 2019;85:164–72.
21. Shen L, Zhao D, Chen Y, Zhang K, Chen X, Lin J, et al. Comparative proteomics analysis of serum proteins in gestational diabetes during early and middle stages of pregnancy. *Proteomics Clin Appl*. 2019;13(5):e1800060.
22. International Association of Diabetes and Pregnancy Study Groups Consensus Panel. International association of diabetes and pregnancy study groups recommendations on the diagnosis and classification of hyperglycemia in pregnancy. *Diabetes Care*. 2010;33(3):676–82. <https://doi.org/10.2337/dc09-1848>.
23. Clinical Laboratory Standards Institute. Procedures for the handling and processing of blood specimens for common laboratory test. 4th ed. Wanye: Clinical Laboratory Standards Institute; 2010.
24. Hou W, Meng X, Zhao W, Pan J, Tang J, Huang Y, et al. Elevated first-trimester Total bile acid is associated with the risk of subsequent gestational diabetes. *Sci Rep*. 2016;6:34070.
25. Xin Y, Hertle E, van der Kallen CJH, Schalkwijk CG, Stehouwer CDA, van Greevenbroek MMJ. Longitudinal associations of the alternative and terminal pathways of complement activation with adiposity: the CODAM study. *Obes Res Clin Pract*. 2018;12(3):286–92.
26. Reynolds R, Hartnett ME, Atkinson JP, Giclas PC, Rosner B, Seddon JM. Plasma complement components and activation fragments: associations with age-related macular degeneration genotypes and phenotypes. *Invest Ophthalmol Vis Sci*. 2009;50(12):5818–27.
27. Kalant D, MacLaren R, Cui W, Samanta R, Monk PN, Laporte SA, et al. CSL2 is a functional receptor for acylation-stimulating protein. *J Biol Chem*. 2005; 280(25):23936–44.
28. Department of Disease Control of Ministry of Health of the People's Republic of China. The guidelines for prevention and control of overweight and obesity in Chinese adults: Beijing: People's Medical Publishing House; 2006. p. 3.
29. Joint Committee for Revising Chinese Guidelines on Prevention and Treatment of Dyslipidemia in Adults. Guidelines on prevention and treatment of dyslipidemia in adults (2016 revision). *Chinese Circulation Journal*. 2016;31(10):937–52.
30. Okesina AB, Donaldson D, Lascelles PT, Morris P. Effect of gestational age on levels of serum alkaline phosphatase isoenzymes in healthy pregnant women. *Int J Gynaecol Obstet*. 1995;48(1):25–9.
31. Bacq Y, Zarka O, Bréchet JF, Mariotte N, Vol S, Tichet J, et al. Liver function tests in normal pregnancy: a prospective study of 103 pregnant women and 103 matched controls. *Hepatology*. 1996;23(5):1030–4.
32. Segers FM, Verdam FJ, de Jonge C, Boonen B, Driessen A, Shiri-Sverdlov R, et al. Complement alternative pathway activation in human nonalcoholic steatohepatitis. *PLoS One*. 2014;9(10):e110053.
33. McCullough RL, McMullen MR, Sheehan MM, Poulsen KL, Roychowdhury S, Chiang DJ, et al. Complement factor D protects mice from ethanol-induced inflammation and liver injury. *Am J Physiol Gastrointest Liver Physiol*. 2018; 315(1):G66–g79.
34. Schmal H, Salzmann GM, Niemeyer P, Langenmair E, Guo R, Schneider C, et al. Early intra-articular complement activation in ankle fractures. *Biomed Res Int*. 2014;2014:426893.
35. Lokki AI, Heikinen-Eloranta J, Jarva H, Saisto T, Lokki ML, Laivuori H, et al. Complement activation and regulation in preeclamptic placenta. *Front Immunol*. 2014;5:312.
36. Basiglio CL, Arriaga SM, Pelusa F, Almará AM, Kapitulnik J, Mottino AD. Complement activation and disease: protective effects of hyperbilirubinaemia. *Clin Sci (Lond)*. 2009;118(2):99–113.
37. Arriaga SM, Mottino AD, Almará AM. Inhibitory effect of bilirubin on complement-mediated hemolysis. *Biochim Biophys Acta*. 1999;1473(2–3): 329–36.
38. Arriaga S, Almará A, Mottino A. In vivo anti-complement effect of bilirubin-IXalpha. *Biochem Pharmacol*. 2002;64(4):741–4.
39. de Paula PF, Barbosa JE, Junior PR, Ferriani VP, Latorre MR, Nudelman V, et al. Ontogeny of complement regulatory proteins - concentrations of factor h, factor i, c4b-binding protein, properdin and vitronectin in healthy children of different ages and in adults. *Scand J Immunol*. 2003;58(5):572–7.
40. Spiga R, Marini MA, Mancuso E, Di Fatta C, Fuoco A, Perticone F, et al. Uric acid is associated with inflammatory biomarkers and induces inflammation via activating the NF-κB signaling pathway in HepG2 cells. *Arterioscler Thromb Vasc Biol*. 2017;37(6):1241–9.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Ready to submit your research? Choose BMC and benefit from:**

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

**At BMC, research is always in progress.**

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

