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Association between plasma leptin and cesarean section after induction of labor: a case control study

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Abstract

Background: Obesity in pregnancy is common, with more than 50% of pregnant women being overweight or obese. Obesity has been identified as an independent predictor of dysfunctional labor and is associated with increased risk of failed induction of labor resulting in cesarean section. Leptin, an adipokine, is secreted from adipose tissue under the control of the obesity gene. Concentrations of leptin increase with increasing percent body fat due to elevated leptin production from the adipose tissue of obese individuals. Interestingly, the placenta is also a major source of leptin production during pregnancy. Leptin has regulatory effects on neuronal tissue, vascular smooth muscle, and nonvascular smooth muscle systems. It has also been demonstrated that leptin has an inhibitory effect on myometrial contractility with both intensity and frequency of contractions decreased. These findings suggest that leptin may play an important role in dysfunctional labor and be associated with the outcome of induction of labor at term. Our aim is to determine whether maternal plasma leptin concentration is indicative of the outcome of induction of labor at term. We hypothesize that elevated maternal plasma leptin levels are associated with a failed term induction of labor resulting in a cesarean delivery.

Methods: In this case-control study, leptin was measured in 3rd trimester plasma samples. To analyze labor outcomes, 174 women were selected based on having undergone an induction of labor (IOL), (115 women with successful IOL and 59 women with a failed IOL). Plasma samples and clinical information were obtained from the UI Maternal Fetal Tissue Bank (IRB# 200910784). Maternal plasma leptin and total protein concentrations were measured using commercially available assays. Bivariate analyses and logistic regression models were constructed using regression identified clinically significant confounding variables. All variables were tested at significance level of 0.05.

Results: Women with failed IOL had higher maternal plasma leptin values (0.5 vs 0.3 pg, $P = 0.01$). These women were more likely to have obesity (mean BMI 32 vs 27 kg/m², $P = 0.0002$) as well as require multiple induction methods (93% vs 73%, $p = 0.008$). Logistic regression showed Bishop score (OR 1.5, $p < 0.001$), BMI (OR 0.92, $P < 0.001$), preeclampsia (OR 0.12, $P = 0.010$), use of multiple methods of induction (OR 0.22, $P = 0.008$) and leptin (OR 0.42, $P = 0.017$) were significantly associated with IOL outcome. Specifically, after controlling for BMI, Bishop Score, and preeclampsia, leptin was still predictive of a failed IOL with an odds ratio of 0.47 ($P = 0.046$). Finally, using leptin as a predictor for

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fetal outcomes, leptin was also associated with of fetal intolerance of labor, with an odds ratio of 2.3 ($P = 0.027$). This association remained but failed to meet statistical significance when controlling for successful (IOL) (OR 1.5, $P = 0.50$).

Conclusions: Maternal plasma leptin may be a useful tool for determining which women are likely to have a failed induction of labor and for counseling women about undertaking an induction of labor versus proceeding with cesarean delivery.

Keywords: Induction of labor, Fetal intolerance of labor, Cesarean section, Vaginal delivery, Leptin, Obesity, Pregnancy

Background

Maternal obesity is associated with a significant increase in the incidence of miscarriage, congenital anomalies, macrosomia, gestational diabetes, gestational hypertension, preeclampsia and stillbirth [1–5]. Obese women have an increased incidence of pre-existing diabetes and chronic hypertension [2] and are more likely to deliver via Cesarean section [2, 3, 5–9]. Obese women are also more likely to undergo IOL [9, 10]. Induction of labor (IOL) is often undertaken because of prevalent co-morbidities such as preeclampsia, hypertension, or diabetes, but obese women are also twice as likely to require induction for postdates pregnancy (> 41–42 weeks gestation) [11–13]. Between 2012 and 2016, 21% of live-births in the United States underwent IOL; of these inductions, 19.2% ended with delivery by Cesarean section [14].

A variety of factors are associated with IOL success, including Bishop score, maternal age and parity [15, 16]. Importantly, obesity has been reported as an independent risk factor for Cesarean delivery following IOL when multiple factors are evaluated, with the highest risk occurring in women with a BMI of 40 or greater [6, 10]. Obesity has also been described as an independent predictor of dysfunctional labor [11, 12, 17–19]. For example, Vahratian et al. showed that labor progression in obese women was significantly slower than normal weight women in early labor [8], and they tended to have a longer total duration of labor and higher oxytocin requirements [9, 20].

Some studies have suggested that dysfunctional labor occurs because there is a reduction in contractility of the obese uterus due to increased cholesterol deposits in the myometrium [19, 21], however, others have found no difference in spontaneous contractile activity of myometrium or response to oxytocin in vitro, attributable to body mass index (BMI) [22]. Leptin, an adipocytokine with an important role in the regulation of energy homeostasis and other neuroendocrine functions [23–26], is found in higher levels in obese individuals, including obese pregnant women [27–31]. Leptin has been shown to be dysregulated in pregnancy, based on maternal condition. For example, individuals suffering from mild and severe preeclampsia had higher levels of leptin compared

to healthy pregnant women [32], where a positive correlation with the levels of the steroid hormone, estradiol was found [33, 34]. Furthermore, leptin is associated with a functional importance during pregnancy due to increasing levels until late second or early third trimester followed by a sharp drop off postpartum [35–37]. Leptin has also been found to exert an inhibitory effect on spontaneous and oxytocin-induced contractions in vitro, with both a decrease in the frequency and amplitude of contractions [37, 38].

Because obese women are at increased risk for failed IOL resulting in a cesarean section, we were interested in the role of leptin in induced labor. More specifically, we and others [39] hypothesize that leptin may play an important role in dysfunctional labor. In this cohort study, we investigate whether elevated plasma leptin levels are associated with the outcome of induction of labor at term.

Methods

Patient selection and plasma collection

Coded samples and clinical data were obtained from the University of Iowa Institutional Review Board (IRB)-approved Maternal-Fetal Tissue Bank (MFTB) [40] (IRB# 200910784). Informed consent was obtained by the MFTB from women to provide biological samples and clinical data from their electronic health record (Epic) for future studies. Women were recruited at prenatal appointments during their first trimester; longitudinal samples and data were collected throughout pregnancy. Samples used in this study were collected between March 2010 and February 2015. The University of Iowa IRB approved this study's protocol (IRB# 201711717) [41]. All local and federal guidelines and assurances were followed. The MFTB provided coded plasma samples and corresponding clinical patient information were obtained for all women who had undergone induction of labor at greater than or equal to 37 weeks gestation and met the following inclusion criteria: induction of labor by any method (misoprostol, dinoprostone, PGE2 gel, oxytocin, artificial rupture of membranes, membrane stripping or Foley balloon placement), complete delivery data in the medical record and availability of plasma sample

collected in the third trimester (>28 weeks). Specific exclusion criteria for this study included multiple gestation, fetal anomalies, augmentation of labor rather than induction of labor, intrauterine fetal demise, and absence of an available plasma sample. Exclusion criteria of the Maternal Fetal Tissue Bank, in general, include HIV+, Hepatitis C+, non-English speaking, inability to provide informed consent, and being less than 18 years of age. Because this study was conducted using previously biobanked samples and corresponding clinical data, no interventions were made in the care of patients. All decisions for medical care including induction and delivery methods were made by the patient and her care team. Blood samples were collected at admission for delivery prior to the beginning of induction. All methods were followed in accordance with this IRB determination and all federal guidelines and assurances.

Blood samples obtained for the MFTB are collected in ACD-A tubes (Becton Dickinson) and are separated into blood plasma and mononuclear cells. Plasma is then aliquoted, snap frozen and stored at -80°C [40]. The MFTB maintains a database to annotate samples with clinical information, which is automatically extracted from the electronic health record and stored in a secure database at the University of Iowa [40]. This database was used to identify women eligible for inclusion in this study. A member of the research team individually verified the clinical information housed in this database for each of the study participants.

From the Maternal Fetal Tissue Bank, all participants that met inclusion/exclusion criteria and had samples available were identified. We were able to retrieve coded 3rd trimester blood plasma samples and clinical data from 174 women who underwent an induction of labor and met inclusion criteria; of these women 59 were delivered via Cesarean section and 115 delivered vaginally.

Coded information on maternal and neonatal characteristics were collected from the MFTB. The data gathered included maternal age at delivery, gestational age at induction, race, parity, body mass index (BMI) (at first presentation to obstetric care), weight gain during pregnancy, indication for induction of labor, method(s) of induction, antepartum/intrapartum complications and characteristics (diabetes, hypertension, preeclampsia, history of Cesarean section, intrauterine growth restriction, Rh isoimmunization, oligohydramnios, chorioamnionitis, presence of meconium-stained fluid, use of epidural/spinal anesthesia during labor), method of delivery (vaginal delivery, vaginal birth after Cesarean [VBAC], operative vaginal delivery, Cesarean section) and indication for Cesarean delivery. Neonatal information including weight, APGAR scores, need for resuscitation at delivery, admission to neonatal intensive care unit,

and presence of respiratory distress, meconium aspiration, or administration of neonatal antibiotic therapy was also collected. The reason for the Cesarean section was determined from the medical record as determined by the provider. Data were collected from the medical record at least 6 weeks postpartum.

Maternal plasma analyses

Leptin levels were measured using the commercially available Human Leptin Instant ELISA (ThermoFisher), which has a sensitivity of 20 pg/mL and a standard curve range of 63–4000 pg/mL. Bicinchoninic acid (BCA) levels were measured using a commercially BCA Protein Assay Kit (ThermoFisher), for the quantitation of total protein, which has a working range of 20–2000 $\mu\text{g/mL}$. All samples were run in duplicate in a single assay. For all analyses, leptin concentration was normalized to micrograms of total protein in the sample and reported as [picogram/total protein [microgram]].

Clinical definitions

Clinical determinations including fetal intolerance of labor, failure to progress, failure to descend, preeclampsia, intrauterine growth restriction, and diabetes were made by the patient's healthcare providers. The determinations made at that time were extracted from the electronic health record (Epic) by the research team.

Statistical analysis

All statistical analyses were performed with SigmaPlot 12.0 software (Systat Software, Inc., California). Logistic regression models were constructed using regression identified and clinically significant confounding variables. In addition, Fisher's exact tests were utilized for categorical variables. For continuous variables, t-test or ANOVA were utilized. Only data available in the electronic health record or from laboratory measurements were used for analyses. All variables were tested at significance level of 0.05. Based on a recent published leptin effect size of 18 pg/mL in control and preeclamptic plasma, for an power of 90% and $\alpha=0.05$ minimally 38 samples per group is necessary [42]. For a parsimonious logistic regression model with 5 covariates minimally 50 samples per group is necessary.

Results

A total of 174 women underwent induction of labor and were included in this study. Characteristics of the study population are summarized in Table 1. Our study included 59 women who ultimately delivered via Cesarean section compared to 115 women who had successful IOL. A successful IOL was defined as having a vaginal delivery, including operative vaginal deliveries with the

Table 1 Patient Demographics, Pregnancy Characteristics, and Leptin Levels

Variable	Successful IOL (vaginal delivery) N = 115	Failed IOL (Cesarean section delivery) N = 59	P Value
Maternal Age at Delivery (mean years, 95% CI)	29.6 (28.6–30.6)	29.9 (28.6–31.2)	0.7
Race: White	87.8% (101)	93.2% (55)	0.7
Race: Black	2.6% (3)	1.7% (1)	0.7
Race: Asian	3.5% (4)	1.7% (1)	0.7
Race: Hispanic	1.7% (2)	0% (0)	0.7
Race: American Indian	0% (0)	1.7% (1)	0.7
Race: Unspecified	3% (3)	1.7% (1)	0.7
Race: Multiracial	1.7% (2)	0% (0)	0.7
BMI (mean kg/m ² , 95% CI)	27.4 (26.2–28.6)	32.2 (29.9–40.0)	0.0002
GA at IOL (mean weeks, 95% CI)	39.8 (39.6–40.0)	39.5 (39.2–39.8)	0.2
Bishop Score (mean, 95% CI)	4.6 (4.1–5.1)	2.6 (2.1–3.1)	<0.001
Parity (mean, 95% CI)	0.9 (0.7–1.1)	0.3 (0.1–0.5)	<0.001
Epidural/Spinal (%), N	84.3% (97)	83.1% (49)	0.9
Weight Gain (mean kg, 95% CI)	12.7 (11.5–13.9)	13.0 (11.4–14.6)	0.8
Induction Method			
Cervidil (dinoprostone) (%), N	28.6% (33)	64.4% (38)	<0.001
Cytotec (misoprostol) (%), N	30.4% (35)	44.1% (26)	0.1
Pitocin (%), N	86.1% (99)	91.5% (54)	0.5
Foley Bulb (%), N	11.3% (13)	27.1% (16)	0.03
AROM (%), N	55.6% (64)	50.8% (30)	0.7
Nipple Stimulation (%), N	0.87% (1)	0%	0.7
Multiple IOL Methods (%), N	73.9% (85)	93.2% (55)	0.008
Indication for IOL			
IOL PET (%), N	1.7% (2)	13.5% (8)	0.008
IOL AMA (%), N	11.3% (13)	18.6% (11)	0.2
IOL Fetal Indication (%), N	18.2% (21)	13.5% (8)	0.5
IOL GDM (%), N	7.8% (9)	3.4% (2)	0.5
IOL DM I (%), N	0.87% (1)	6.8% (4)	0.2
IOL DM II (%), N	0% (0)	1.7% (1)	0.7
IOL CHTN (%), N	11.3% (13)	15.2% (9)	0.6
IOL gHTN (%), N	8.6% (10)	3.4% (2)	0.4
IOL Postdates (%), N	28.6% (33)	30.5% (18)	0.9
IOL PROM (%), N	6.1% (7)	6.8% (4)	0.8
IOL Elective (%), N	25.2% (29)	8.5% (5)	0.03
IOL Multiple Indications (%), N	19.1% (22)	18.6% (11)	0.9
Antepartum/Intrapartum Complications			
No Pregnancy Complications (%), N	45.2% (52)	30.5% (18)	0.06
CHTN (%), N	11.3% (13)	15.2% (9)	0.06
gHTN (%), N	8.6% (10)	3.4% (2)	0.4
Preeclampsia (%), N	1.7% (2)	13.5% (8)	0.008
DM I (%), N	0.8% (1)	6.7% (4)	0.07
DM II (%), N	0% (0)	1.7% (1)	0.2
GDM A1 (%), N	4.3% (5)	3.4% (2)	0.7
GDM A2 (%), N	6.1% (7)	3.4% (2)	0.7
Alloimmunization (%), N	0.8% (1)	1.7% (1)	0.8
IUGR (%), N	1.7% (2)	0% (0)	0.8
Oligohydramnios (%), N	7.8% (9)	1.7% (1)	0.2
Cerclage (%), N	2.6% (3)	1.7% (1)	0.8

Table 1 (continued)

Variable	Successful IOL (vaginal delivery) N = 115	Failed IOL (Cesarean section delivery) N = 59	P Value
Hypothyroid (%), N)	11.3% (13)	8.5% (5)	0.9
AMA (%), N)	12.2% (14)	18.6% (11)	0.4
History of Cesarean Section (%), N)	4.3% (5)	1.7% (1)	0.7
VBAC (%), N)	1.7% (2)	0% (0)	0.8
Operative Vaginal Delivery (%), N)	14.8% (17)	0% (0)	0.007
Meconium (%), N)	13.0% (15)	35.6% (21)	0.002
Chorioamnionitis (%), N)	7.8% (9)	13.6% (8)	0.4
Gestational age at sample collection	39 5/7 weeks	38 5/7 weeks	0.26
Leptin (pg/ug)	0.3 (0.3–0.4)	0.5 (0.4–0.7)	0.01

BMI body mass index, GA gestational age, IOL induction of labor, AROM artificial rupture of membranes, AMA advanced maternal age (>= 35 years), GDM gestational onset diabetes, DM I type I diabetes mellitus, DM II type II diabetes mellitus, CHTN chronic hypertension, gHTN gestational hypertension, PROM premature rupture of membranes, GDM A1 gestational onset diabetes mellitus type I, GDM A2 gestational onset diabetes mellitus type II, IUGR intrauterine growth restriction, VBAC vaginal birth after Cesarean

Data are presented as mean with 95% confidence interval or percentage with N. Categorical variables were compared using Chi square. Continuous variables were analyzed using t-Test or ANOVA. α = 0.05

assistance of either forceps or vacuum as this was the most frequent utilized definition in a systematic review related to induction of labor. At baseline, the two groups were comparable in terms of maternal age, race, parity, gestational age at induction, weight gain during pregnancy, and use of an epidural during labor. Women with Cesarean section were more likely to have obesity as defined by BMI ≥ 30 kg/m² at the beginning of pregnancy (mean BMI 32 vs 27 kg/m², P = 0.0002), preeclampsia (13% vs 2%, P = 0.008), lower Bishop score (3 vs 5, P < 0.001), lower parity (0.3 vs 0.9, P < 0.001) and meconium-stained fluid (35% vs 13%, P = 0.002). Method(s) of induction of labor in women ultimately delivered via Cesarean section, was/were more likely to include dinoprostone (65% vs 29%, P < 0.001) and Foley balloon placement (26% vs 11%, P = 0.03). They were also more likely to require multiple induction methods as defined by use of any two or more methods (93% vs 73%, P = 0.008).

The indication for Cesarean delivery was failed operative delivery in which the use of forceps and/or vacuum preceded a cesarean delivery (2%), arrest of dilation (failure to progress) (26%), arrest of descent (failure to descend) (31%), and fetal intolerance of labor (52%). Percentages do not equal 100 because the indication for Cesarean section may have included one or more of the above diagnoses (i.e. failure to progress and fetal intolerance of labor).

Neonatal characteristics are summarized in Table 2. Women with Cesarean section were more likely to have infants with lower APGAR scores at 1 and 5 min (6 vs 8, P < 0.001 and 8 vs 9, P = 0.008, respectfully), higher need for resuscitation at time of delivery (54% vs 15%, P < 0.001), as well as the diagnoses of respiratory distress syndrome (31% vs 4%, P < 0.001) and meconium aspiration syndrome (9% vs 0.9%, P = 0.02).

Maternal plasma leptin levels in women with Cesarean section were higher than those who had a vaginal delivery

Table 2 Neonatal Characteristics

Neonatal Characteristic	Successful IOL	Failed IOL	P Value
Birth Weight (mean grams, 95% CI)	3494 (3395–3593)	3645 (3504–3785)	0.05
APGAR 1 min (mean, 95% CI)	7.9 (7.6–8.2)	6.3 (5.7–6.9)	< 0.001
APGAR 5 min (mean, 95% CI)	8.8 (8.7–8.9)	8.4 (8.1–8.7)	0.008
Resuscitation (%), N)	14.8% (17)	54.2% (32)	< 0.001
Respiratory Distress (%), N)	4.3% (5)	30.5% (18)	< 0.001
Neonatal Antibiotics (%), N)	15.6% (18)	22.0% (13)	0.5
Meconium Aspiration (%), N)	0.86% (1)	8.5% (5)	0.02
NICU Stay (Mean Days, 95% CI)	1.0 (0–2.2)	1.2 (0.5–1.9)	0.8

NICU Neonatal Intensive Care Unit

Data are presented as mean with 95% confidence interval or percentage with N. Categorical variables were compared using Chi square. Continuous variables were analyzed using t-Test or ANOVA. α = 0.05

(0.5 vs 0.3 leptin pg/ug, $P=0.01$). Figure 1 demonstrates the association between leptin and successful IOL versus Cesarean section delivery, with a 2 tailed Student's t test $P=0.01$.

A regression analysis was performed, and the results of the various models are shown in Table 3. Bishop score (OR 1.5, $P<0.001$), BMI (OR 0.92, $P<0.001$), preeclampsia (0.12, $P=0.01$), use of multiple methods of induction (OR 0.22, $P=0.008$) and leptin (OR 0.42, $P=0.02$) were the only covariates significantly associated with successful IOL. BMI and leptin were co-linear variables, as shown in Fig. 2. Thus, after controlling for Bishop score and preeclampsia, leptin was still predictive of successful IOL with an odds ratio of 0.47 ($P=0.046$). Finally, a model using leptin as a predictor for fetal outcomes demonstrated that leptin was also predictive of fetal intolerance of labor (FIOL), with an odds ratio of 2.3 ($P=0.03$). These models are shown in Table 4 and Fig. 3.

Discussion

The pathophysiology of obesity in pregnancy is not completely understood but involves a host of environmental and genetic factors. A complex physiologic system is responsible for the regulation of energy homeostasis and the adipocytokine, leptin, is among the key players [23]. Leptin is a 16-kDa polypeptide product of the *LEP* gene (also known as *OB*, *OBS*, *LEPD*) in humans, first isolated by Zhang et al. [43] in 1994 and mapped to chromosome 7 in 1995 [44], although an obesity causing mutation in mice was described long before [45, 46]. Leptin is produced primarily by adipocytes and its secretion is highly proportional to body fat content [23, 28, 30, 47]; and more specifically, to the stored amount of lipid in the fat cells of adipose tissue [29]. Obese individuals have been demonstrated to have markedly elevated leptin levels

Table 3 Logistic Regression Models and Association with Successful Induction of Labor

Model	Bishop Score	BMI	Leptin	Preeclampsia	Multiple IOL Methods
1	1.5 (<0.001)				
2		0.92 (<0.001)			
3			0.42 (0.017)		
4				0.12 (0.010)	
5					0.22 (0.008)
6	1.5 (<0.001)	0.93 (0.004)			
7	1.5 (<0.001)		0.49 (0.056)		
8	1.4 (<0.001)	0.94 (0.004)		0.186 (0.046)	
9	1.4 (<0.001)		0.47 (0.046)	0.16 (0.037)	
10	1.4 (<0.001)	0.94 (0.008)		0.22 (0.071)	0.28 (0.06)
11	1.4 (<0.001)		0.45 (0.057)	0.20 (0.058)	0.25 (0.043)

Data are presented as odds ratio with p value for each independent variable. The dependent variable for each model is the occurrence of a successful induction of labor

[27–30]. As one of the primary signals from the body's energy stores, leptin acts on the central nervous system (CNS) to inhibit appetite and promote satiety and energy expenditure [23, 25]. Because obese individuals have high circulating levels, the concept of leptin resistance

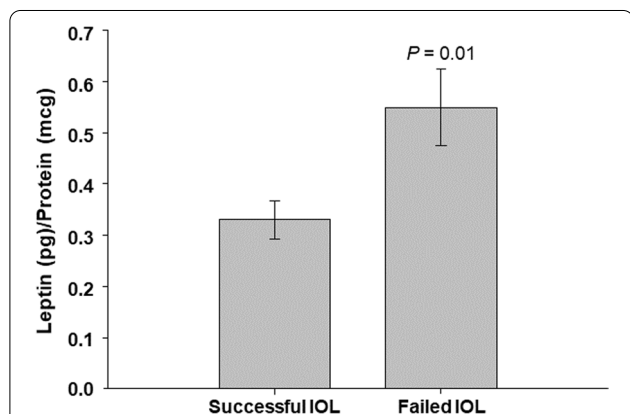


Fig. 1 Leptin and successful IOL versus cesarean delivery. Leptin is significantly higher in women with failed IOL compared to successful IOL using a Student's two-tailed t test ($P=0.01$)

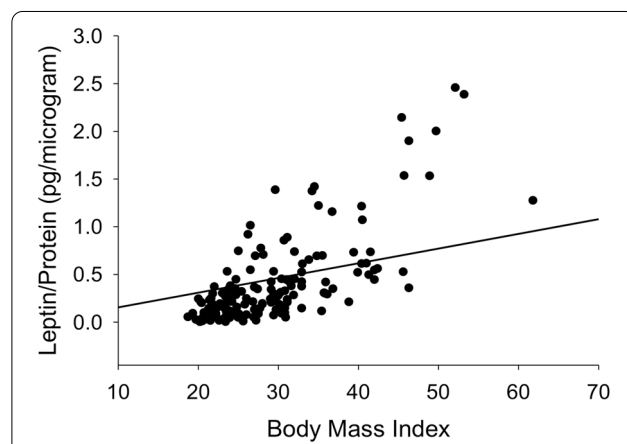
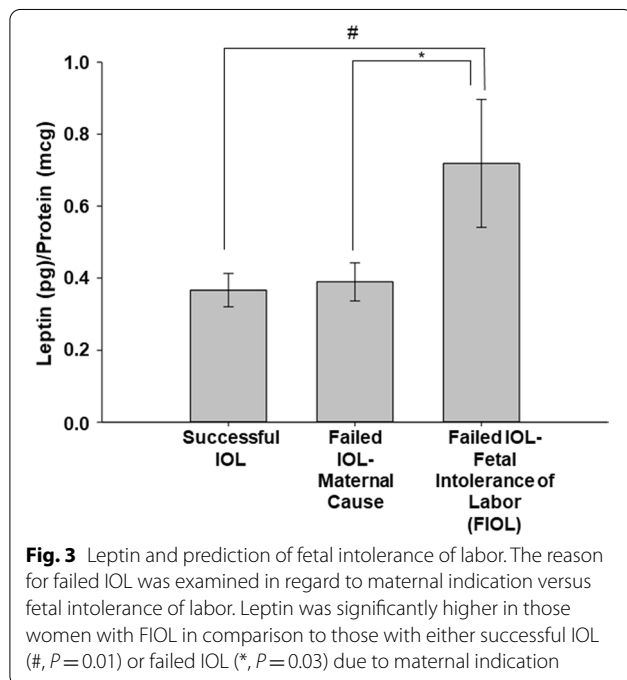


Fig. 2 Correlation between Leptin and BMI. We and others demonstrate that leptin and BMI are highly correlated and therefore, cannot be used in the same model ($R^2=0.73$, $P<0.001$)

Table 4 Linear regression models and association with fetal outcomes

Model	Apgar 1	Respiratory Distress	NICU Days	Fetal Intolerance of Labor (FIOL)	Successful IOL
1	0.80 (0.486)				
2		0.88 (0.850)			
3			0.34 (0.254)		
4				2.3 (0.027)	
5				1.5 (0.50)	1.0 (<0.0001)

Data are presented as odds ratio with *p* value for each independent variable. The dependent variable for each model is Leptin level



emerged [24, 25]. Characterized by elevated serum leptin levels and decreased leptin sensitivity, leptin resistance is not completely understood but may represent a fundamental pathology of obesity.

To our knowledge, this is the first-time maternal 3rd trimester leptin has been shown to be associated with failed induction of labor resulting in cesarean section, with an odds ratio of 0.47 ($P=0.046$). Not surprisingly, leptin levels were associated with increasing BMI. This is consistent with prior data that has shown maternal levels in pregnancy increase linearly with pre-pregnancy BMI

[31]. Thus, in future studies to better elucidate the role of leptin, the Cesarean section and vaginal delivery after IOL and groups will need to be better matched for BMI. We hypothesize that increased circulating leptin levels play an important role in the pathophysiology of dysfunctional labor due to an inhibitory effect on uterine smooth muscle, thus significantly increasing the likelihood for Cesarean section delivery. As previously highlighted, prior research has shown that leptin exerts an inhibitory effect on spontaneous and oxytocin-induced myometrial contractions, with both a decrease in the frequency and amplitude of contractions [38]. Leptin also has the ability to inhibit myometrial apoptosis [48] and extracellular matrix remodeling [49], both proposed as essential steps needed for the uterus to develop powerful and synchronous contractions during labor [50–54]. In addition, extracellular matrix remodeling has been implicated in rupture of membranes and cervical ripening [55, 56]. Our results support the concept of a possible metabolic regulation of the human myometrium during pregnancy and labor and highlight one of the proposed mechanisms for dysfunctional labor and failed induction of labor in obese women.

Obesity is well-established as a significant predictor of failed IOL, but induction is often medically indicated for any one of the varieties of pregnancy-related complications that are more common in obese women. The risk for an emergency or unplanned Cesarean delivery in nulliparous women increases proportionally with a patient’s BMI [7] and although the increased morbidity associated with Cesarean delivery following prolonged labor or rupture of membranes is true for all women, the obese population are at greater risk for complications in these settings [57, 58], including higher rates of infection, wound complications, and postpartum hemorrhage [2]. The risk of failed induction and increased rate of complications with cesarean delivery puts the clinician in a difficult position when counseling about induction of labor. Thus, research aimed at identifying factors that are associated with the outcome of induction of labor is clinically important.

Currently, the Bishop score has traditionally been one of the main factors used to predict success of labor induction. The Bishop score assesses position, consistency, effacement and dilatation of the cervix and the fetal station [59]. However, a systematic review of 40 articles found the Bishop score to be a poor predictor [60]. This may be due to the subjective nature of the measurement that can vary between observers [61]. Thus, there continues to be a gap in being able to counsel pregnant women about the possibility of successful induction of labor, especially in the setting of obesity. Additionally, having a better prediction of the

mode of delivery can help healthcare providers determine the best healthcare facility for delivery. This can be especially helpful in settings in which there is not 24-h onsite access to a surgeon or anesthesia. To fill these gaps, researchers need to identify a reproducible and objective method to predict cesarean delivery [62]. Objective methods for predicting successful induction of labor may include the use of ultrasound or biomarkers.

Strengths of the study were adequate power to detect a clinically significant difference in outcomes. Also, using the Maternal-Fetal Tissue Bank provides a regulated, maintained, and unbiased platform for clinical research, with consistency established in the acquisition and storage of tissue/plasma samples and clinical information. The consistency in sample collection and handling is critical to biomarker studies. Weaknesses of our study include the study's retrospective design and relatively homogenous study population which may limit generalizability, but should not bias the measurements of leptin.

Conclusions

In order for prenatal care providers to be able to identify which patients are ideal candidates for a successful induction of labor, there needs to be a better understanding of the physiological mechanisms involved in this process. Because leptin can suppress uterine contractility, it is not surprising that elevated leptin levels are associated with failed induction. Taken together, this suggests a strong mechanistic role for leptin in the regulation of labor which we are continuing to investigate.

Abbreviations

IOL: Induction of labor; IRB: Institutional Review Board; MFTB: Maternal Fetal Tissue Bank; BMI: Body mass index; FIOL: Fetal intolerance of labor; VBAC: Vaginal birth after cesarean section; pg: Picograms; mL: Milliliter; BCA: Bicinchoninic acid assay; ug: Microgram; GA: Gestational age; AROM: Artificial rupture of membranes; AMA: Advanced maternal age; GDM: Gestational onset diabetes mellitus; DM1: Type I Diabetes Mellitus; DMII: Type II Diabetes Mellitus; CHTN: Chronic hypertension; gHTN: Gestational hypertension; PROM: Premature rupture of membranes; GDM A1: Gestational onset diabetes mellitus Type I; GDM A2: Gestational onset diabetes mellitus Type II; IUGR: Intrauterine growth restriction; NICU: Neonatal intensive care unit.

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None.

Authors' contributions

WC designed and performed experiments, performed data collection and analysis, and in writing the manuscript. SS assisted in experimental design, data analysis, and in writing the manuscript. WH processed samples and assisted in data collection. AK assisted in data analysis and manuscript preparation. NB assisted in experiment design, data analysis, and in writing the manuscript. ED participated in data collection and analysis and in writing the manuscript. MS participated in experimental design, data analysis, and manuscript preparation. DS assisted in designing and performing experiments, data analysis, and manuscript preparation. All authors read and approved the final transcript.

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Availability of data and materials

The data that support the findings of this study are available from the Maternal Fetal Tissue Bank at the University of Iowa but restrictions apply to the availability of these data, which were used under an IRB-approved agreement for the current study, and so are not publicly available. Data are however available from the authors upon reasonable request and with permission of the Maternal Fetal Tissue Bank. Requests for data and/or samples should be made to Donna Santillan, PhD (corresponding author).

Declarations

Ethics approval and consent to participate

Informed consent was obtained from participants by the Maternal Fetal Tissue Bank at the University of Iowa (IRB#200910784). The University of Iowa IRB approved this study's protocol (IRB# 201711717). All local and federal guidelines and assurances were followed.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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References

1. American College of Obstetricians and Gynecologists. ACOG Committee opinion no. 549: obesity in pregnancy. *Obstet Gynecol.* 2013;121(1):213–7.
2. Fyfe EM, Anderson NH, North RA, Chan EH, Taylor RS, Dekker GA, et al. Screening for pregnancy endpoints C: risk of first-stage and second-stage cesarean delivery by maternal body mass index among nulliparous women in labor at term. *Obstet Gynecol.* 2011;117(6):1315–22.
3. Robinson HE, O'Connell CM, Joseph KS, McLeod NL. Maternal outcomes in pregnancies complicated by obesity. *Obstet Gynecol.* 2005;106(6):1357–64.
4. Weiss JL, Malone FD, Emig D, Ball RH, Nyberg DA, Comstock CH, et al. Obesity, obstetric complications and cesarean delivery rate—a population-based screening study. *Am J Obstet Gynecol.* 2004;190(4):1091–7.
5. Howell KR, Powell TL. Effects of maternal obesity on placental function and fetal development. *Reproduction.* 2017;153(3):R97–R108.
6. Chu SY, Kim SY, Schmid CH, Dietz PM, Callaghan WM, Lau J, et al. Maternal obesity and risk of cesarean delivery: a meta-analysis. *Obes Rev.* 2007;8(5):385–94.
7. Poobalan AS, Aucott LS, Gurung T, Smith WC, Bhattacharya S. Obesity as an independent risk factor for elective and emergency caesarean delivery in nulliparous women—systematic review and meta-analysis of cohort studies. *Obes Rev.* 2009;10(1):28–35.
8. Vahratian A, Siega-Riz AM, Savitz DA, Zhang J. Maternal pre-pregnancy overweight and obesity and the risk of cesarean delivery in nulliparous women. *Ann Epidemiol.* 2005;15(7):467–74.
9. Ellis JA, Brown CM, Barger B, Carlson NS. Influence of maternal obesity on labor induction: a systematic review and Meta-analysis. *J Midwifery Womens Health.* 2019;64(1):55–67.
10. Wolfe KB, Rossi RA, Warshak CR. The effect of maternal obesity on the rate of failed induction of labor. *Am J Obstet Gynecol.* 2011;205(2):128 e121–7.

11. Arrowsmith S, Wray S, Quenby S. Maternal obesity and labour complications following induction of labour in prolonged pregnancy. *BJOG*. 2011;118(5):578–88.
12. Denison FC, Price J, Graham C, Wild S, Liston WA. Maternal obesity, length of gestation, risk of postdates pregnancy and spontaneous onset of labour at term. *BJOG*. 2008;115(6):720–5.
13. Roos N, Sahlin L, Ekman-Ordeberg G, Kieler H, Stephansson O. Maternal risk factors for postterm pregnancy and cesarean delivery following labor induction. *Acta Obstet Gynecol Scand*. 2010;89(8):1003–10.
14. Rossi RM, Requarth E, Warshak CR, Dufendach KR, Hall ES, DeFranco EA. Risk calculator to predict cesarean delivery among women undergoing induction of labor. *Obstet Gynecol*. 2020;135(3):559–68.
15. Vrouwenraets FP, Roumen FJ, Dehing CJ, van den Akker ES, Aarts MJ, Scheve EJ. Bishop score and risk of cesarean delivery after induction of labor in nulliparous women. *Obstet Gynecol*. 2005;105(4):690–7.
16. Yeast JD, Jones A, Poskin M. Induction of labor and the relationship to cesarean delivery: a review of 7001 consecutive inductions. *Am J Obstet Gynecol*. 1999;180(3 Pt 1):628–33.
17. Ovesen P, Rasmussen S, Kesmodel U. Effect of prepregnancy maternal overweight and obesity on pregnancy outcome. *Obstet Gynecol*. 2011;118(2 Pt 1):305–12.
18. Shirazian T, Raghavan S. Obesity and pregnancy: implications and management strategies for providers. *Mt Sinai J Med*. 2009;76(6):539–45.
19. Zhang J, Bricker L, Wray S, Quenby S. Poor uterine contractility in obese women. *BJOG*. 2007;114(3):343–8.
20. Pevzner L, Powers BL, Rayburn WF, Rumney P, Wing DA. Effects of maternal obesity on duration and outcomes of prostaglandin cervical ripening and labor induction. *Obstet Gynecol*. 2009;114(6):1315–21.
21. Smith RD, Babychuk EB, Noble K, Draeger A, Wray S. Increased cholesterol decreases uterine activity: functional effects of cholesterol alteration in pregnant rat myometrium. *Am J Phys Cell Phys*. 2005;288(5):C982–8.
22. Higgins CA, Martin W, Anderson L, Blanks AM, Norman JE, McConnachie A, et al. Maternal obesity and its relationship with spontaneous and oxytocin-induced contractility of human myometrium in vitro. *Reprod Sci*. 2010;17(2):177–85.
23. Budak E, Fernandez Sanchez M, Bellver J, Cervero A, Simon C, Pellicer A. Interactions of the hormones leptin, ghrelin, adiponectin, resistin, and PYY3-36 with the reproductive system. *Fertil Steril*. 2006;85(6):1563–81.
24. Kaur J. A comprehensive review on metabolic syndrome. *Cardiol Res Pract*. 2014;2014:943162.
25. Pan H, Guo J, Su Z. Advances in understanding the interrelations between leptin resistance and obesity. *Physiol Behav*. 2014;130:157–69.
26. Wroblewski A, Strycharz J, Swiderska E, Drewniak K, Drzewoski J, Szmraj J, et al. Molecular insight into the interaction between epigenetics and leptin in metabolic disorders. *Nutrients*. 2019;11(8):1872.
27. Hamilton BS, Paglia D, Kwan AY, Deitel M. Increased obese mRNA expression in omental fat cells from massively obese humans. *Nat Med*. 1995;1(9):953–6.
28. Korner J, Leibel RL. To eat or not to eat - how the gut talks to the brain. *N Engl J Med*. 2003;349(10):926–8.
29. Lonnqvist F, Nordfors L, Jansson M, Thorne A, Schalling M, Arner P. Leptin secretion from adipose tissue in women. Relationship to plasma levels and gene expression. *J Clin Invest*. 1997;99(10):2398–404.
30. Maffei M, Halaas J, Ravussin E, Pratley RE, Lee GH, Zhang Y, et al. Leptin levels in human and rodent: measurement of plasma leptin and Ob RNA in obese and weight-reduced subjects. *Nat Med*. 1995;1(11):1155–61.
31. Tsai PJ, Davis J, Bryant-Greenwood G. Systemic and placental leptin and its receptors in pregnancies associated with obesity. *Reprod Sci*. 2015;22(2):189–97.
32. Gutaj P, Sibiak R, Jankowski M, Awdi K, Bryl R, Mozdzia P, et al. The Role of the Adipokines in the Most Common Gestational Complications. *Int J Mol Sci*. 2020;21(24):9408.
33. Atamer Y, Erden AC, Demir B, Kocyigit Y, Atamer A. The relationship between plasma levels of leptin and androgen in healthy and preeclamptic pregnant women. *Acta Obstet Gynecol Scand*. 2004;83(5):425–30.
34. Henson MC, Castracane VD. Leptin in pregnancy: an update. *Biol Reprod*. 2006;74(2):218–29.
35. Tessier DR, Ferraro ZM, Gruslin A. Role of leptin in pregnancy: consequences of maternal obesity. *Placenta*. 2013;34(3):205–11.
36. Perez-Perez A, Toro A, Vilarino-Garcia T, Maymo J, Guadix P, Duenas JL, et al. Leptin action in normal and pathological pregnancies. *J Cell Mol Med*. 2018;22(2):716–27.
37. Hajagos-Toth J, Ducza E, Samavati R, Vari SG, Gaspar R. Obesity in pregnancy: a novel concept on the roles of adipokines in uterine contractility. *Croat Med J*. 2017;58(2):96–104.
38. Moynihan AT, Hehir MP, Glavey SV, Smith TJ, Morrison JJ. Inhibitory effect of leptin on human uterine contractility in vitro. *Am J Obstet Gynecol*. 2006;195(2):504–9.
39. Wuntakal R, Kaler M, Hollingworth T. Women with high BMI: should they be managed differently due to antagonising action of leptin in labour? *Med Hypotheses*. 2013;80(6):767–8.
40. Santillan MK, Leslie KK, Hamilton WS, Boese BJ, Ahuja M, Hunter SK, et al. Collection of a lifetime: a practical approach to developing a longitudinal collection of women's healthcare biological samples. *Eur J Obstet Gynecol Reprod Biol*. 2014;179:94–9.
41. Coded private information or specimens use in research, Guidance (2008) [<https://www.hhs.gov/ohrp/regulations-and-policy/guidance/research-involving-coded-private-information/index.html#>].
42. Bawah AT, Yeboah FA, Nanga S, Alidu H, Ngala RA. Serum adipocytokines and adiposity as predictive indices of preeclampsia. *Clin Hypertens*. 2020;26:19.
43. Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature*. 1994;372(6505):425–32.
44. Green ED, Maffei M, Braden VV, Proenca R, DeSilva U, Zhang Y, et al. The human obese (OB) gene: RNA expression pattern and mapping on the physical, cytogenetic, and genetic maps of chromosome 7. *Genome Res*. 1995;5(1):5–12.
45. Ingalls AM, Dickie MM, Snell GD. Obese, a new mutation in the house mouse. *J Hered*. 1950;41(12):317–8.
46. Leibel RL, Bahary N, Friedman JM. Genetic variation and nutrition in obesity: approaches to the molecular genetics of obesity. *World Rev Nutr Diet*. 1990;63:90–101.
47. Considine RV, Sinha MK, Heiman ML, Kriauciunas A, Stephens TW, Nyce MR, et al. Serum immunoreactive-leptin concentrations in normal-weight and obese humans. *N Engl J Med*. 1996;334(5):292–5.
48. Wendremaire M, Bardou M, Peyronel C, Hadi T, Sagot P, Morrison JJ, et al. Effects of leptin on lipopolysaccharide-induced myometrial apoptosis in an in vitro human model of chorioamnionitis. *Am J Obstet Gynecol*. 2011;205(4):363 e361–9.
49. Wendremaire M, Mourtilion P, Goirand F, Lirussi F, Barrichon M, Hadi T, et al. Effects of leptin on lipopolysaccharide-induced remodeling in an in vitro model of human myometrial inflammation. *Biol Reprod*. 2013;88(2):45.
50. Charpigny G, Leroy MJ, Breuille-Fouche M, Tanfin Z, Mhaouty-Kodja S, Robin P, et al. A functional genomic study to identify differential gene expression in the preterm and term human myometrium. *Biol Reprod*. 2003;68(6):2289–96.
51. Leong AS, Norman JE, Smith R. Vascular and myometrial changes in the human uterus at term. *Reprod Sci*. 2008;15(1):59–65.
52. Monga M, Sanborn BM. Uterine contractile activity. *Introduction. Semin Perinatol*. 1995;19(1):1–2.
53. Shynlova O, Oldenhof A, Dorogin A, Xu Q, Mu J, Nashman N, et al. Myometrial apoptosis: activation of the caspase cascade in the pregnant rat myometrium at midgestation. *Biol Reprod*. 2006;74(5):839–49.
54. Shynlova O, Tsui P, Jaffer S, Lye SJ. Integration of endocrine and mechanical signals in the regulation of myometrial functions during pregnancy and labour. *Eur J Obstet Gynecol Reprod Biol*. 2009;144(Suppl 1):S2–10.
55. Stygar D, Wang H, Vlastic YS, Ekman G, Eriksson H, Sahlin L. Increased level of matrix metalloproteinases 2 and 9 in the ripening process of the human cervix. *Biol Reprod*. 2002;67(3):889–94.
56. Yonemoto H, Young CB, Ross JT, Guilbert LL, Fairclough RJ, Olson DM. Changes in matrix metalloproteinase (MMP)-2 and MMP-9 in the fetal amnion and chorion during gestation and at term and preterm labor. *Placenta*. 2006;27(6–7):669–77.
57. Allen VM, O'Connell CM, Baskett TF. Maternal morbidity associated with cesarean delivery without labor compared with induction of labor at term. *Obstet Gynecol*. 2006;108(2):286–94.

58. Goffman D, Madden RC, Harrison EA, Merkatz IR, Chazotte C. Predictors of maternal mortality and near-miss maternal morbidity. *J Perinatol.* 2007;27(10):597–601.
59. Bishop EH. Pelvic scoring for elective induction. *Obstet Gynecol.* 1964;24:266–8.
60. Kolkman DG, Verhoeven CJ, Brinkhorst SJ, van der Post JA, Pajkrt E, Opmeer BC, et al. The Bishop score as a predictor of labor induction success: a systematic review. *Am J Perinatol.* 2013;30(8):625–30.
61. Faltin-Traub EF, Boulvain M, Faltin DL, Extermann P, Irion O. Reliability of the Bishop score before labour induction at term. *Eur J Obstet Gynecol Reprod Biol.* 2004;112(2):178–81.
62. Kamel RA, Negm SM, Youssef A, Bianchini L, Brunelli E, Pilu G, et al. Predicting cesarean delivery for failure to progress as an outcome of labor induction in term singleton pregnancy. *Am J Obstet Gynecol.* 2021;224(6):609 e601–11.

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